

The formation of adrenaline from noradrenaline*

It has been demonstrated that the methyl group of methionine can be utilized *in vivo* for the formation of adrenaline¹. Similarly, adrenaline-¹⁴C has been isolated from the adrenal glands of the rat after injection of noradrenaline-¹⁴C². The formation of adrenaline in homogenates of dog adrenal glands has been reported³, but attempts to repeat this work gave negative results².

BORSOOK AND DUBNOFF⁴ have shown that ATP and methionine are required for the methylation of guanidinoacetic acid; CANTONI⁵ has demonstrated that S-adenosylmethionine (AMe) is the active methylating agent. The methylation of noradrenaline to form adrenaline by an enzyme system obtained from bovine adrenal medullae which utilizes either ATP and methionine, or S-adenosylmethionine, is reported here.

After the incubations described below, 1 μ mole of unlabeled adrenaline was added to each of the incubation mixtures, and trichloroacetic acid extracts were then prepared. Adrenaline and noradrenaline were separated from each other by ion-exchange chromatography⁶ and the radioactivity in the adrenaline fractions was determined with a thin window gas flow counter with an average background of 16 c.p.m.

L-Methionine-methyl-¹⁴C was obtained from the Volk Radio-Chemical Company. S-Adenosylmethionine-methyl-¹⁴C was prepared as described by CANTONI⁷.

Table I shows the distribution of enzyme activity in homogenates of fresh adrenal medullae. The medullae were homogenized in ice cold 0.25 *M* sucrose. After removing the unbroken cells by centrifuging at $1000 \times g$ for 10 min, the homogenate was centrifuged at $20,000 \times g$ for 30 min. The supernatant was decanted and saved for assay. The sediment was washed twice in 0.25 *M* sucrose and finally suspended in a volume of 0.25 *M* sucrose to equal the volume of the $20,000 \times g$ supernatant fraction. It can be seen that with either ATP and methionine, or with S-adenosylmethionine as substrates, essentially all of the enzyme activity is found in the supernatant fraction.

TABLE I
ADRENALINE FORMATION IN ADRENAL MEDULLARY CELL FRACTIONS

Cell fraction	Methyl donor	Adrenaline c.p.m.
1. 0.5 ml supernatant + 0.5 ml particles	Methionine	1140
2. 0.5 ml particles	Methionine	40
3. 0.5 ml supernatant	Methionine	1600
4. 0.5 ml particles	S-Adenosylmethionine	70
5. 0.5 ml supernatant	S-Adenosylmethionine	3530

To each reaction mixture was added MgCl_2 , 200 μ moles; reduced glutathione, 2.5 mg; *l*-noradrenaline, 1 μ mole; and 0.2 ml tris buffer, pH 7.4, 0.5 *M*. Nos. 1, 2 and 3 contained 10 μ moles ATP and 1 μ mole L-methionine-methyl-¹⁴C ($3.1 \cdot 10^5$ c.p.m.). Nos. 4 and 5 contained 0.36 μ mole S-adenosylmethionine-methyl-¹⁴C ($5 \cdot 10^4$ c.p.m.). The total volume was 1.8 ml. Incubations were for 2 h at 37° under nitrogen.

Table II presents data on the $(\text{NH}_4)_2\text{SO}_4$ fractionation of the enzymes. Adrenal medullae were homogenized in 2.5 volumes of cold 0.1 *M* sodium acetate, pH 5.6. The homogenate was centrifuged at $20,000 \times g$ for 30 min. The supernatant was decanted through glass wool and fractionated by adding the calculated volumes of saturated $(\text{NH}_4)_2\text{SO}_4$ solution. Prior to use, each of the enzyme preparations was dialyzed against 0.1 *M* sodium acetate, pH 6.6, for 4 h. All operations were performed at 2°. Both the methionine activating enzyme (MAE) and noradrenaline methyltransferase (NMP) are found in the 0.35 to 0.50 saturated $(\text{NH}_4)_2\text{SO}_4$ fractions.

Comparing the amounts of radioactivity in the adrenaline fractions when ATP and methionine are used as co-substrates to the amounts found when AMe is used as co-substrate, and considering that the initial amount of radioactivity in the AMe was one tenth of the amount of radioactivity in the methionine, it is seen that AMe is utilized at about 20 to 30 times the rate of ATP and methionine for the methylation of noradrenaline.

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TABLE II
AMMONIUM SULFATE FRACTIONATION OF NORADRENALINE METHYLPHERASE

	% (NH ₄) ₂ SO ₄ saturation	C.p.m./μmole adrenaline/mg protein		% Total activity	
		ATP + Methionine	AME	ATP + Methionine	AME
1.	0	1220	2540	100	100
2.	0-35	410	1110	7	8
3.	35-50	1610	4640	31	38
4.	50-75	745	2040	4	5

Each reaction mixture contained reduced glutathione, 2.5 mg; MgCl₂, 200 μmoles; *l*-noradrenaline, 1 μmole; 0.2 ml tris buffer 0.5 *M*, pH 7.5; and either 0.14 μmole S-adenosylmethionine-methyl-¹⁴C (2.7 · 10⁴ c.p.m.) or 10 μmoles ATP and 1.0 μmole L-methionine-methyl-¹⁴C (3.1 · 10⁵ c.p.m.). Each incubated 2 h at 37° under nitrogen. 1, 2, 3 and 4 contained, respectively, 2.0, 2.8, 3.0, and 2.3 mg of protein/ml; the total volume in each instance was 1.0 ml.

Table III shows the absolute requirement for ATP and Mg⁺⁺ when methionine is used as the methyl donor. CANTONI⁵ has reported these requirements for the MAE and also reported a requirement for glutathione or other -SH compounds for optimal activity of MAE. When AME is the methyl donor, glutathione is required for optimal activity; Mg⁺⁺ also appears to be necessary. CANTONI AND VIGNOS⁸ reported a requirement for -SH compounds for optimal activity of guanidinoacetate methyltransferase, but Mg⁺⁺ was not required.

TABLE III
REQUIREMENTS FOR ADRENALINE FORMATION

Reaction medium	C.p.m./μmole adrenaline/mg protein	
	Methionine	AME
Complete system	1610	4640
No glutathione	1510	2870
No Mg ⁺⁺	0	3370
No ATP	0	
No noradrenaline	200	170

The complete system contained MgCl₂, 200 μmoles; reduced glutathione 2.5 mg; *l*-noradrenaline, 1 μmole; 0.2 ml tris buffer, pH 7.4, 0.5 *M*; ATP, 10 μmoles when methionine was used as co-substrate; and either 1 μmole L-methionine-methyl-¹⁴C (3.1 · 10⁵ c.p.m.) or 0.14 μmole S-adenosylmethionine-methyl-¹⁴C (2.7 · 10⁴ c.p.m.). Final volume was 1 ml. The protein concentration was 3.0 mg/ml. Incubated 2 h at 37° under nitrogen.

NORMAN KIRSHNER
McC. GOODALL*

Departments of Biochemistry and Physiology,
Duke University School of Medicine, Durham, N.C. (U.S.A.)

¹ E. B. KELLER, R. A. BOISSONAS AND V. DU VIGNEAUD, *J. Biol. Chem.*, 183 (1950) 627.

² D. T. MASUOKO, H. F. SCHOTT, R. I. AKAWIE AND W. G. CLARK, *Proc. Soc. Exptl. Biol. Med.*, 93 (1956) 5.

³ E. BULLBRING, *Brit. J. Pharmacol.*, 4 (1949) 234.

⁴ H. BORSOOK AND J. W. DUBNOFF, *J. Biol. Chem.*, 171 (1947) 363.

⁵ G. L. CANTONI, *J. Biol. Chem.*, 204 (1953) 403.

⁶ N. KIRSHNER AND McC. GOODALL, *J. Biol. Chem.*, 226 (1957) (in the press).

⁷ G. L. CANTONI, in S. P. COLOWICK AND N. O. KAPLAN, *Methods in Enzymology*, Vol. III, Academic Press Inc., New York, 1957, p. 600.

⁸ G. L. CANTONI AND P. J. VIGNOS, JR., *J. Biol. Chem.*, 209 (1954) 647.

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